



Análisis SARS-CoV-2 en Aguas Residuales

Viruses in human feces

Pneumotropic

Influenza
SARS CoV-1
SARS-CoV-2
MERS-CoV

Multitropic

Ebola



Neurotropic

Enterovirus

Poliovirus

Parechovirus

Nipah virus

TBE

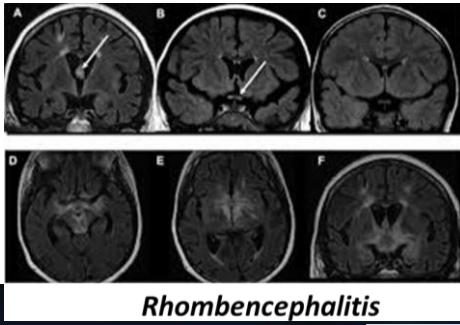
Hepatotropic

Hepatitis A
Hepatitis E

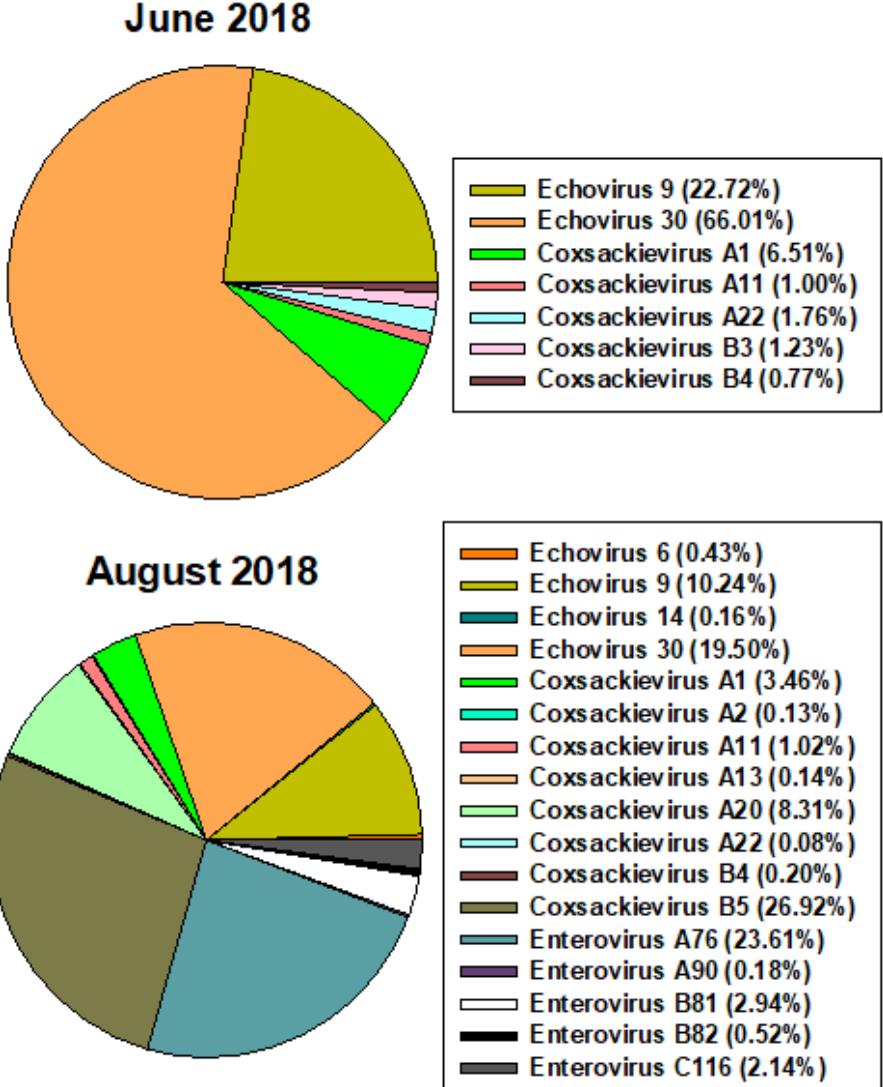
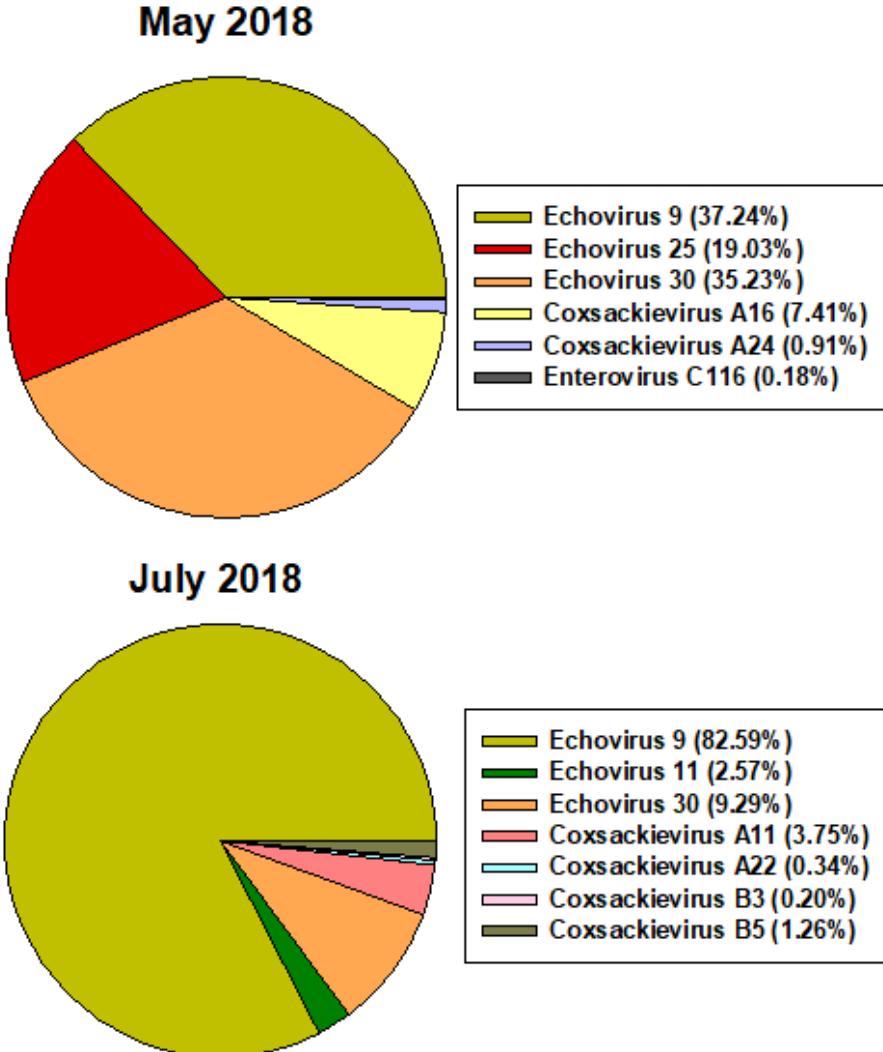
Enterotropic

Norovirus
Sapovirus
Rotavirus
Astrovirus
Adenovirus
Aichivirus





Enteroviruses in Barcelona Raw Sewage





EL CORONAVIRUS SARS-CoV-2 Y LA PANDEMIA DE COVID-19



Sociedad Española de Virología (SEV)

25/4/2020

Manejo clínico del paciente de COVID-19

- La mayor parte de los pacientes (80-85%) tienen una enfermedad leve y sin complicaciones [ver Ficha #InfoSEV nº 8]
- Algunos (15-20%) desarrollan cuadros clínicos mas graves, que requieren hospitalización y oxigenoterapia.
- Aproximadamente un 5% del total de infectados requieren ingreso en la unidad de cuidados intensivos (UCI).
- En el punto de urgencias: valoración inmediata del riesgo de cada paciente; aislamiento y uso de mascarilla por el paciente; personal sanitario con equipo de protección adecuado.
- Para saber más: <https://www.who.int/docs/default-source/coronavirus/clinical-management-of-novel-cov.pdf>

Cuadro clínico	Síntomas	Medidas
Síntomas leves	Fiebre Dolor de cabeza Fatiga, dolor muscular Diarrea, anorexia, vómitos	Antipiréticos, hidratación Paracetamol Reposo Tto. sintomático, vigilancia Ingreso en hospital, oxigenoterapia, antivirales, anti-inflamatorios
Neumonía severa	Respiración rápida, letargia, saturación O ₂ baja Co-infecciones Síntomas neurológicos	Antibioterapia específica Prevención complicaciones, Inmunosupresores Ventilación en pronación UCI: Intubación, ventilación mecánica
Síndrome de distress respiratorio	Dificultad respiratoria Dificultad respiratoria severa	
Sepsis	Problemas de coagulación, síntomas neurológicos, alteraciones urinarias	Heparina, tto. específico
Shock séptico	Hipotensión, taquicardia, taquipnea	Tto. específico



<https://www.mscbs.gob.es/>

<https://www.isciii.es/>

<http://sevirologia.es/>



@sanidadgob

@CIBER_ISCIII
@sev_virologia

Cite as: M. M. Lamers *et al.*,
Science 10.1126/science.abc1669 (2020).

SARS-CoV-2 productively infects human gut enterocytes

Mart M. Lamers^{1*}, Joep Beumer^{2*}, Jelte van der Vaart^{2*}, Kèvin Knoops³, Jens Puschkhof², Tim I. Breugem¹, Raimond B. G. Ravelli³, J. Paul van Schayck³, Anna Z. Mykytyn¹, Hans Q. Duimel³, Elly van Donselaar³, Samra Riesebosch¹, Helma J. H. Kuijpers³, Debby Schippers¹, Willine J. van de Wetering³, Miranda de Graaf¹, Marion Koopmans¹, Edwin Cuppen^{4,5}, Peter J. Peters³, Bart L. Haagmans^{1†}, Hans Clevers^{2†‡}



picture alliance/dpa

Christian Drosten (La Charité, Berlin):
The proportion of infectious SARS-CoV-2 in respiratory secretions is very low (around 1 infectious unit in 10^7 physical particles) and even less in feces

A close-up, high-magnification microscopic image showing numerous small, red, irregularly shaped objects. These represent individual SARS-CoV-2 viral particles, likely viewed under a fluorescence microscope to highlight specific markers on the virus.

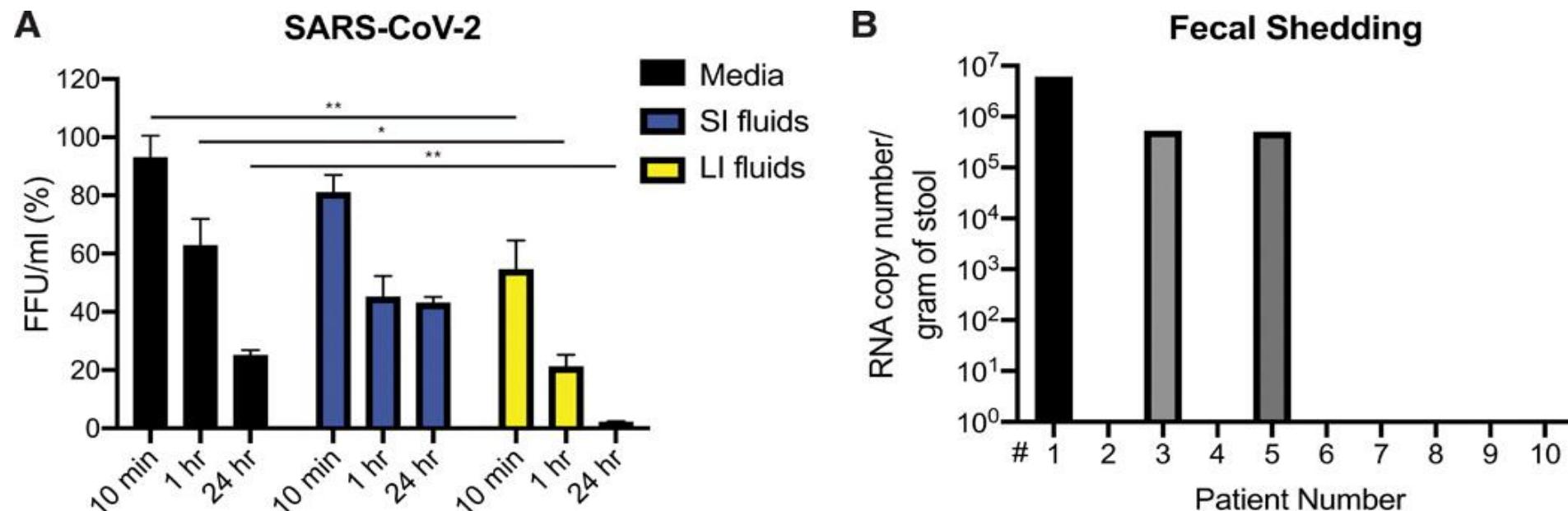
Cite as: R. Zang *et al.*, *Sci. Immunol.* 10.1126/sciimmunol.abc3582 (2020).

CORONAVIRUS

TMPRSS2 and TMPRSS4 promote SARS-CoV-2 infection of human small intestinal enterocytes

Ruochen Zang^{1,2,*}, Maria Florencia Gomez Castro^{1,*}, Broc T. McCune³, Qiru Zeng¹, Paul W. Rothlauf^{1,4}, Naomi M. Sonnek⁵, Zhuoming Liu¹, Kevin F. Brulois^{6,7}, Xin Wang², Harry B. Greenberg^{7,8}, Michael S. Diamond^{1,3,9}, Matthew A. Ciorba⁵, Sean P. J. Whelan¹, Siyuan Ding^{1†}

SARS-CoV-2 rapidly lose infectivity in the human GI tract.



Disclaimer: Early release articles are not considered as final versions. Any changes will be reflected in the online version in the month the article is officially published.

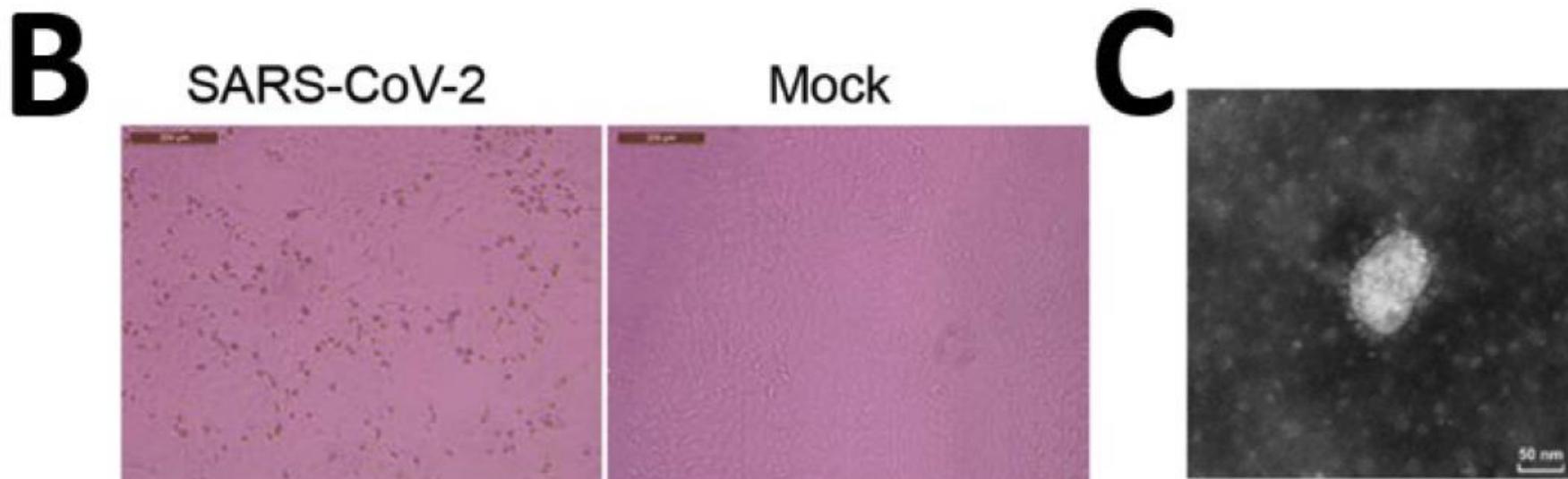
Volume 26, Number 8—August 2020

Research Letter

Infectious SARS-CoV-2 in Feces of Patient with Severe COVID-19

Fei Xiao¹, Jing Sun¹, Yonghao Xu¹, Fang Li¹, Xiaofang Huang¹, Heying Li, Jingxian Zhao, Jicheng Huang, and Jincun Zhao✉

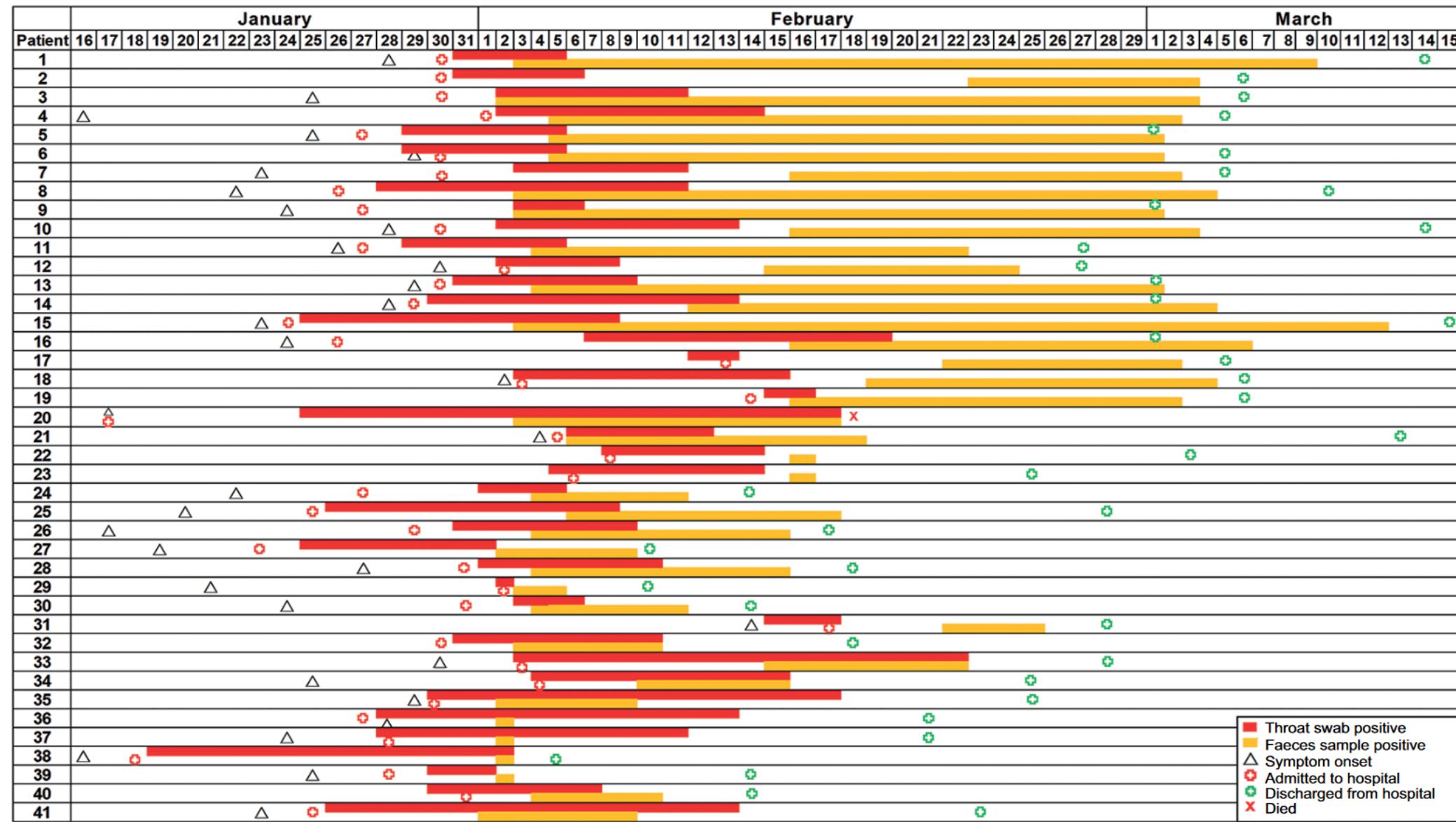
Author affiliations: Sun Yat-sen University, Zhuhai, China (F. Xiao); Guangzhou Medical University, Guangzhou, China (J. Sun, Y. Xu, F. Li, X. Huang, Jingxian Zhao, Jincun Zhao); Chinese Academy of Sciences, Guangzhou (H. Li); Guangzhou Customs District Technology Center, Guangzhou (J. Huang)



B) Vero E6 cells infected with SARS-CoV-2 isolate for 72 hours. C) Detection of viral particles by using transmission electron microscopy (original magnification, $\times 98,000$).

Prolonged Presence of SARS-CoV-2 Viral RNA in Faecal Samples

Wu et al., 2020 The Lancet, [https://doi.org/10.1016/S2468-1253\(20\)30083-2](https://doi.org/10.1016/S2468-1253(20)30083-2)



Timeline of results from throat swabs and faecal samples through the course of disease



- Medema G, Heijnen L, Elsinga G, Italiaander R, Brouwer A. Presence of SARS-CoV-2 in sewage. *medRxiv*. 2020.03.29.20045880
- Lodder W, de Roda Husman AM. SARS-CoV-2 in wastewater: potential health risk, but also data source. *The Lancet Gastroenterology & Hepatology*. 2020;5(6):533-4.

Water Research 181 (2020) 115942

Contents lists available at ScienceDirect

Water Research

journal homepage: www.elsevier.com/locate/watres

 ELSEVIER



SARS-CoV-2 RNA in wastewater anticipated COVID-19 occurrence in a low prevalence area

Walter Randazzo ^{a,b}, Pilar Truchado ^c, Enric Cuevas-Ferrando ^b, Pedro Simón ^d, Ana Allende ^c, Gloria Sánchez ^{b,*}

^a Department of Microbiology and Ecology, University of Valencia, Av. Dr. Moliner, 50, Burjassot, 46100, Valencia, Spain
^b Department of Preservation and Food Safety Technologies, Institute of Agrochemistry and Food Technology, IATA-CSIC, Av. Agustín Escardino 7, Paterna, 46980, Valencia, Spain
^c Research Group on Quality, Safety and Bioactivity of Plant Foods, Department of Food Science and Technology, CEBAS-CSIC, Campus Universitario de Espinardo, 25, 30100, Murcia, Spain
^d ESAMUR, Avenida Juan Carlos, s/n - Edificio Torre Jemeca, Murcia, Spain

Sentinel surveillance of SARS-CoV-2 in wastewater anticipates the occurrence of COVID-19 cases

Gemma Chavarria-Miró, Eduard Anfruns-Estrada,  Susana Guix, Miquel Paraira, Belén Galofré, Gloria Sánchez,  Rosa Pintó,  Albert Bosch

doi: <https://doi.org/10.1101/2020.06.13.20129627>

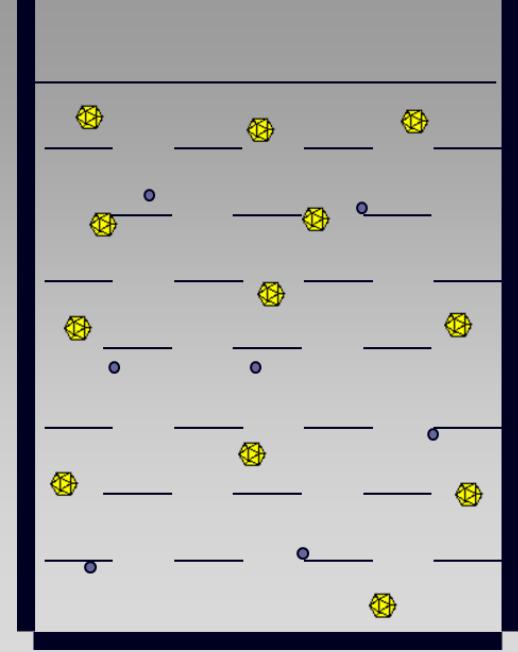
medRxiv
THE PREPRINT SERVER FOR HEALTH SCIENCES



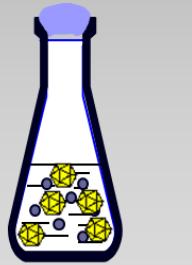
BMJ Yale



Virus detection in the water environment:



A problem that
requires
concentration...



Food and Environmental Virology (2019) 11:184–192
<https://doi.org/10.1007/s12560-019-09378-0>

ORIGINAL PAPER



Glass Wool Concentration Optimization for the Detection of Enveloped and Non-enveloped Waterborne Viruses

Albert Blanco^{1,2} · Islem Abid³ · Nawal Al-Otaibi³ · Francisco José Pérez-Rodríguez^{1,2} · Cristina Fuentes^{1,2} ·
Susana Guix^{1,2} · Rosa M. Pintó^{1,2} · Albert Bosch^{1,2}

PROTOCOLO DETECCIÓN SARS-CoV-2 EN AGUA RESIDUAL

REACTIVOS

- Tiras reactivas de pH
- Tampón TGEB pH 9.5 (100 mM Tris –HCl, Glicina 0.05M, Extracto de carne 1%)
- PEG 6000
- NaCl
- NaOH 5M
- HCl 5M
- PBS pH 7.4
- Kit de extracción de RNA (NucliSENS® miniMAG® extraction system, BioMérieux)
- RNA UltraSense™ One-Step Quantitative RT-PCR System, Invitrogen)
- Virus control de proceso: Transmissible Gastroenteritis Enteric Virus (TGEV) [1]
- Twist Synthetic SARS-CoV-2 RNA Control 2 (MN908947.3) (Twist Bioscience)

PROTOCOLO DETECCIÓN DE SARS-CoV-2 EN AGUAS RESIDUALES (Versión 1.4, Junio 2020)

ENVÍO DE LAS MUESTRAS

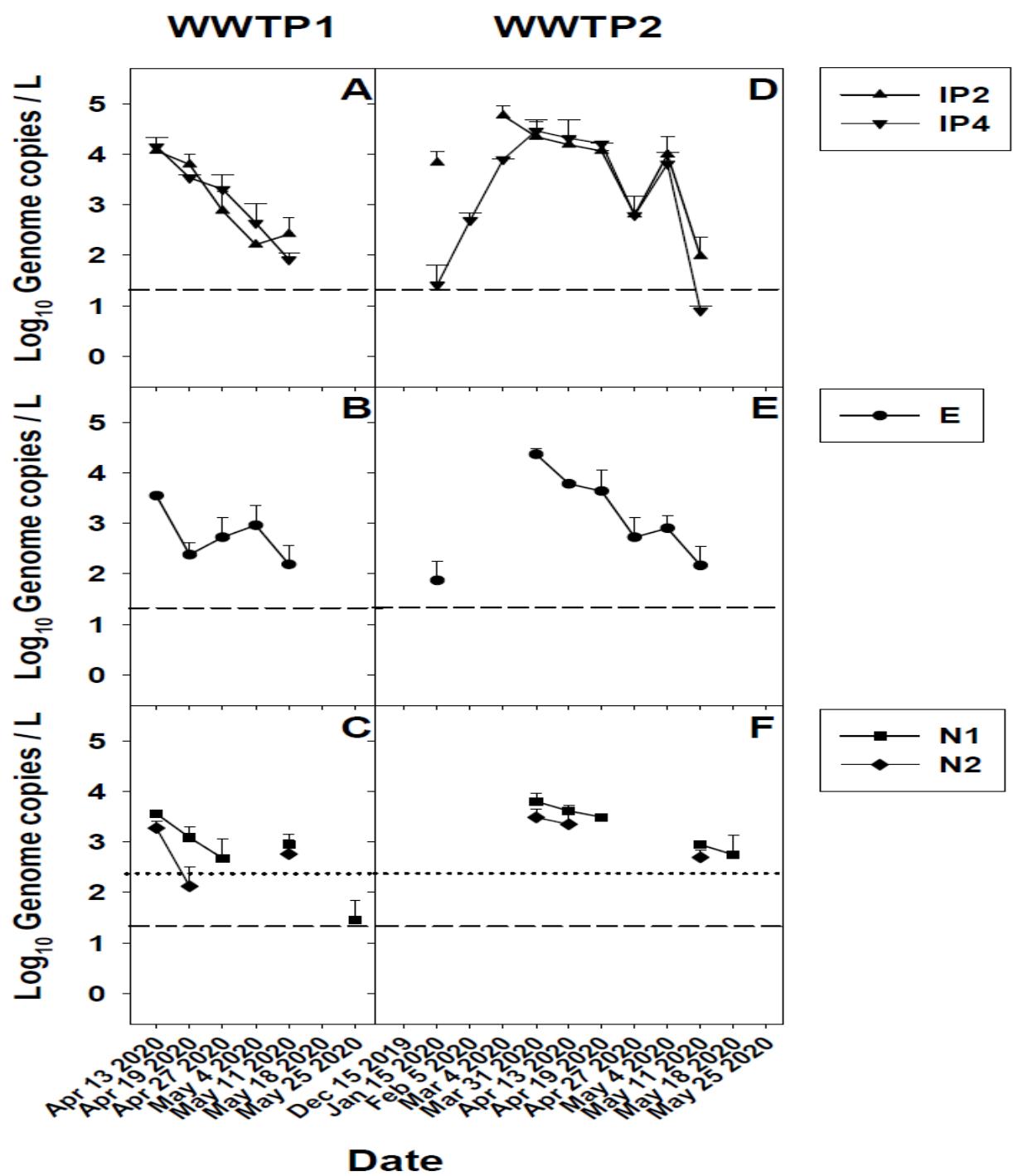
- El envío de las muestras de aguas desde la EDAR hasta los laboratorios de análisis se debe de realizar mediante transporte refrigerado.
- Las botellas de las muestras de agua se deben introducir en cajas herméticas que impidan el derrame en caso de rotura. Dentro de las cajas se debe introducir algún material absorbente que evite derrames en caso de rotura.





- **EDAR Besòs: 3 M inhabitant equivalents**
- **EDAR El Prat de Llobregat: 2 M inhabitant equivalents**





Primer regions:

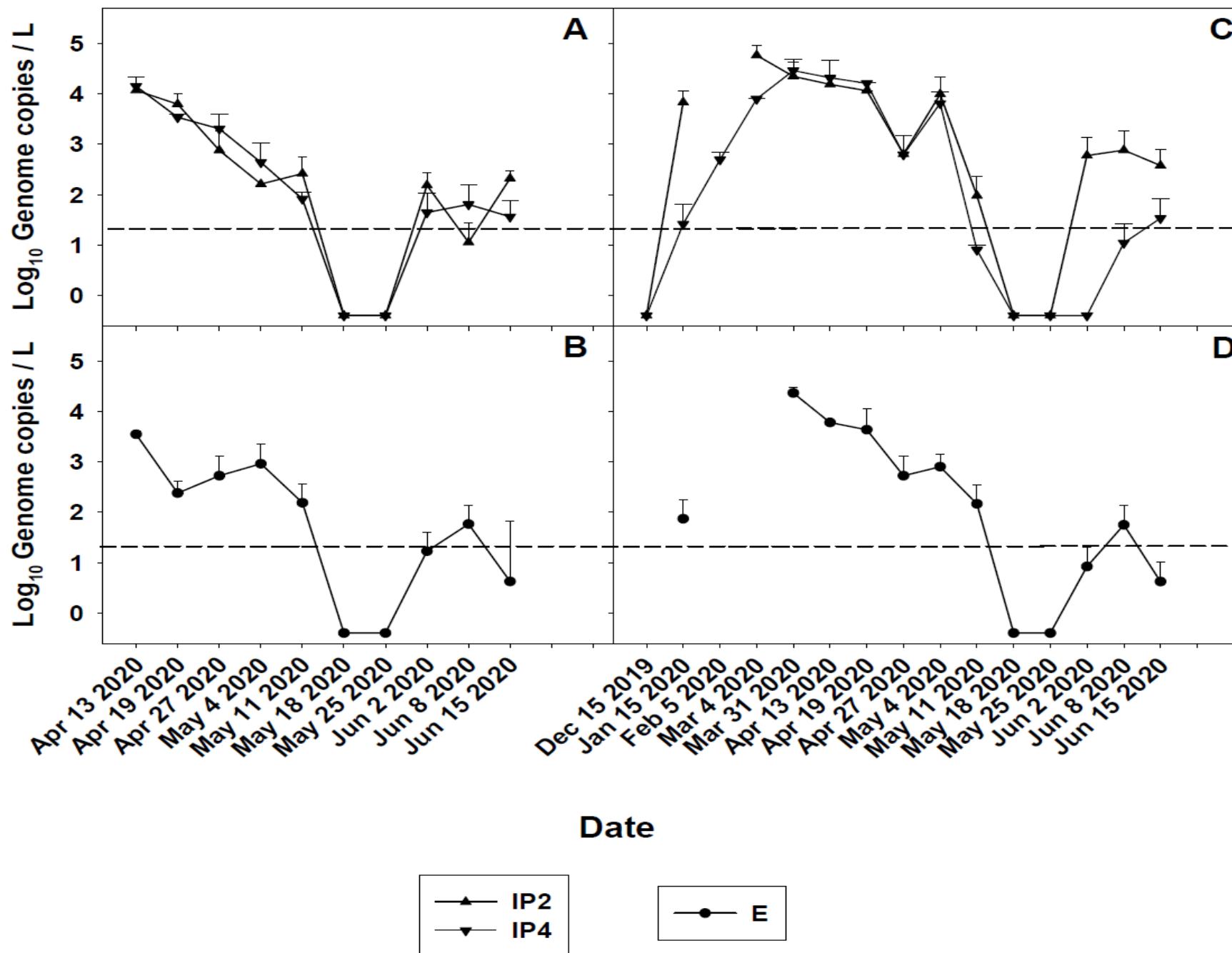
RdRp: **IP2, IP4**, Institut Pasteur

Envelope protein: **E**, Charité Berlin

Nucleoprotein: **N1, N2**, CDC

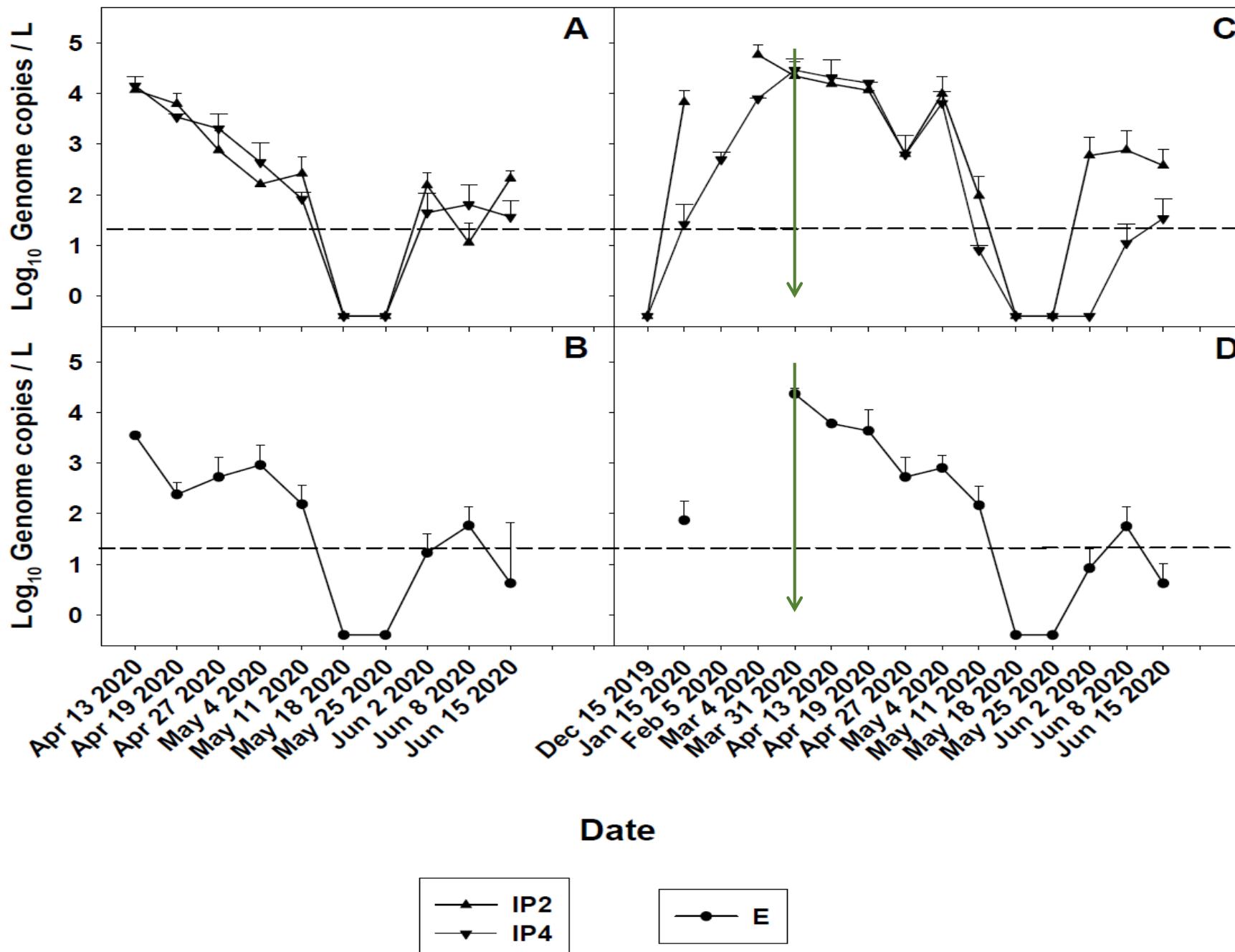
WWTP1 (Besòs)

WWTP2 (El Prat)



WWTP1 (Besòs)

WWTP2 (El Prat)



Primer muestreo: 31 de marzo

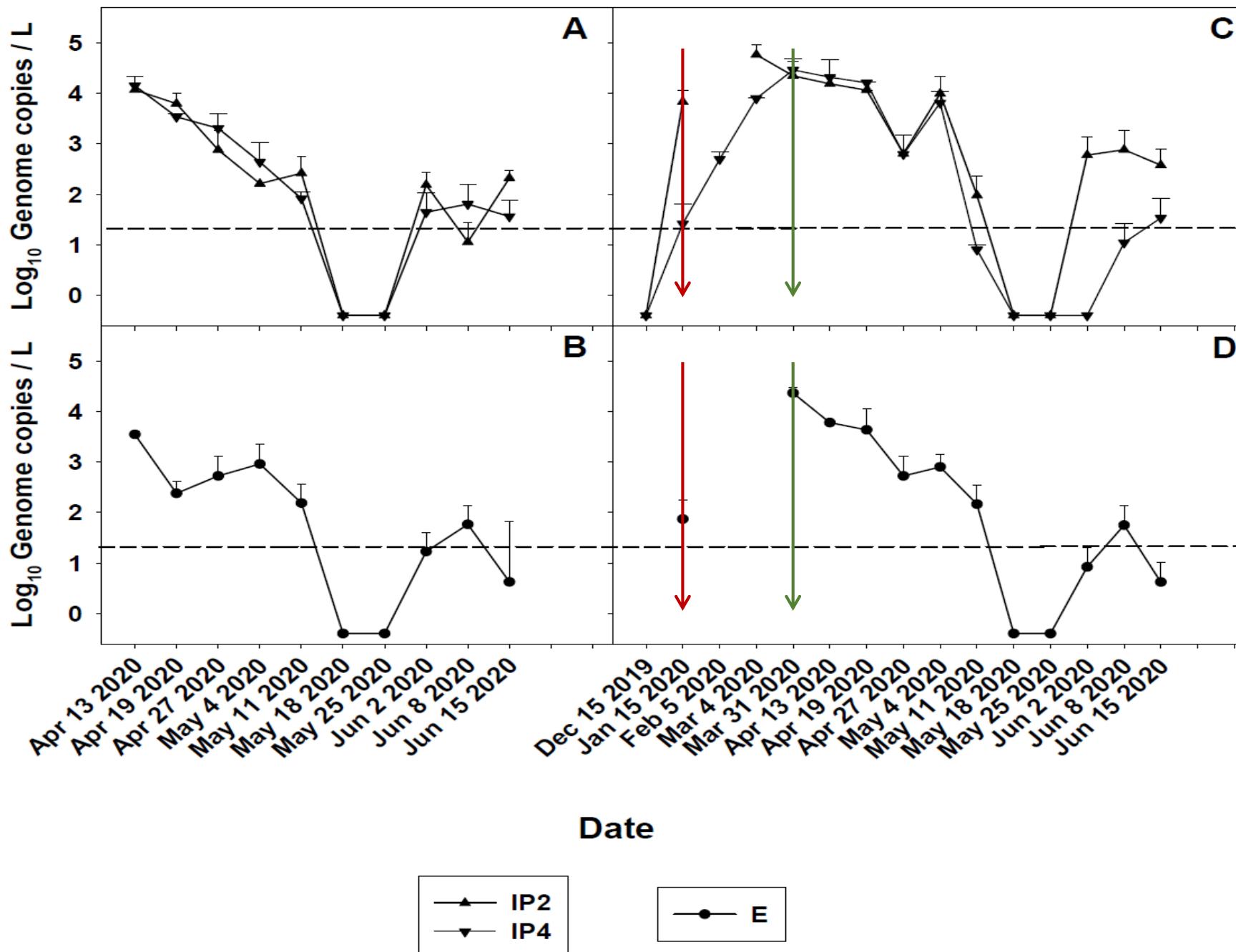


VIRUS ENTÈRICS



WWTP1 (Besòs)

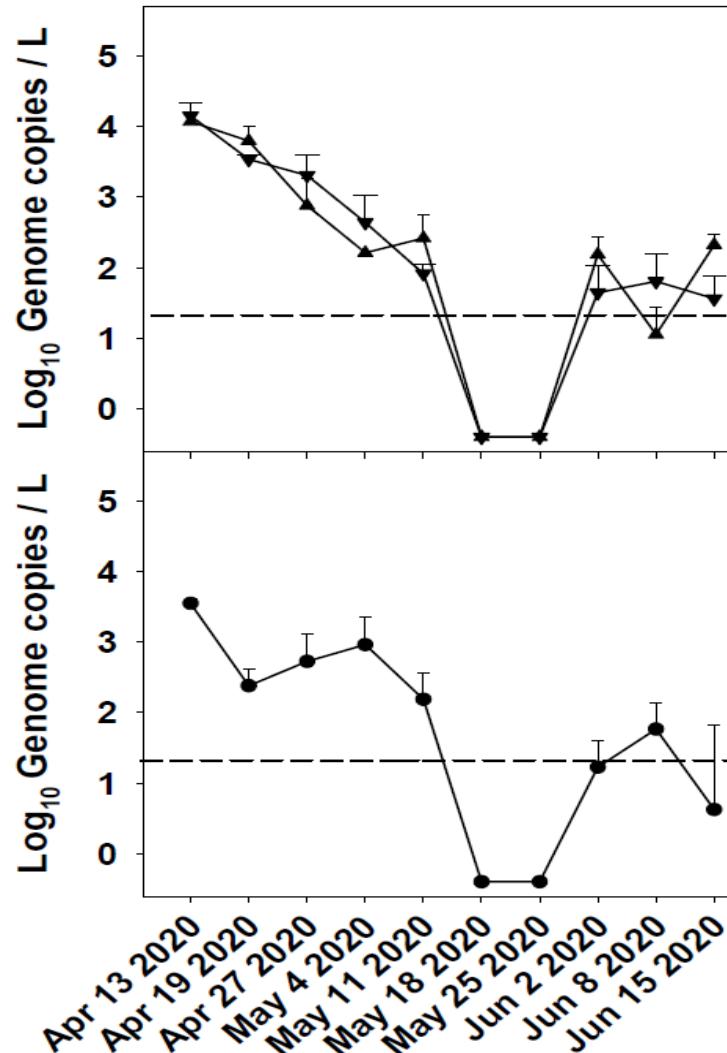
WWTP2 (El Prat)



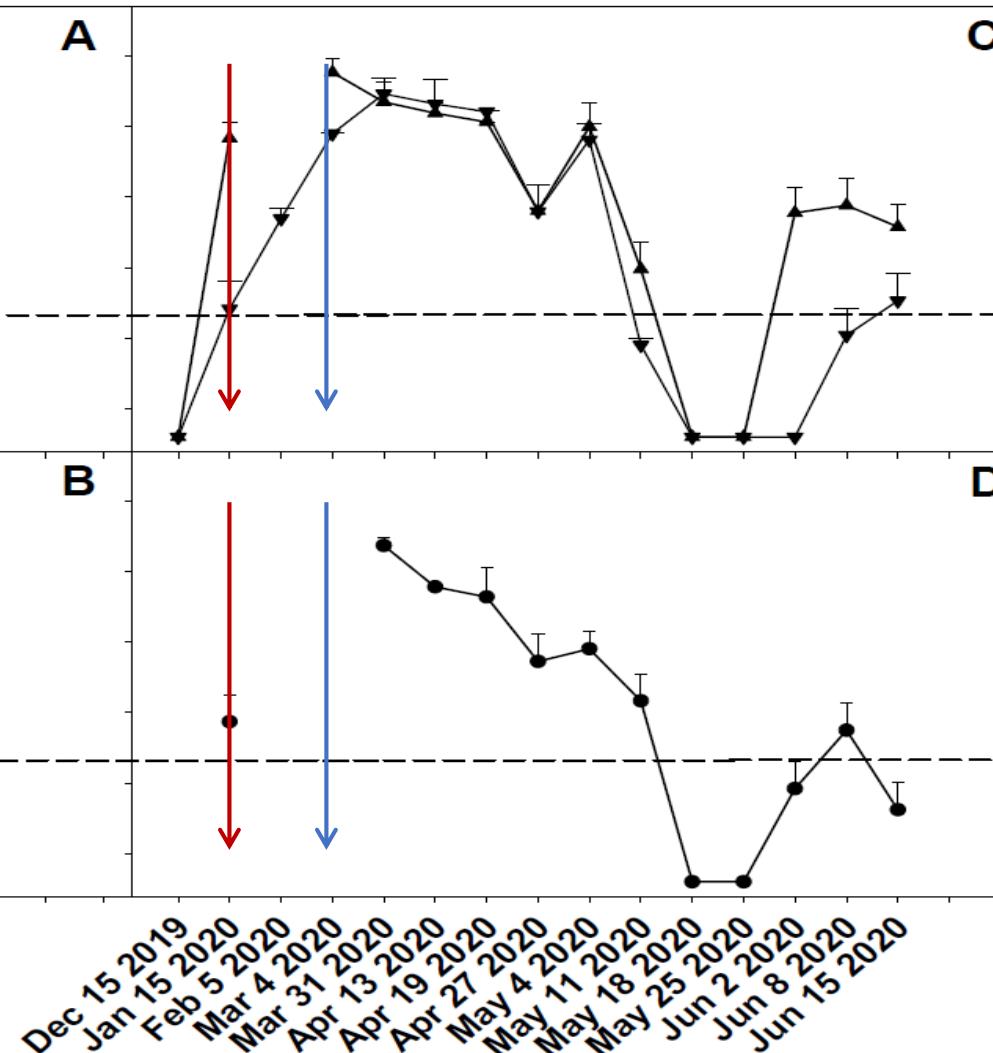
Primer muestreo: 31 de marzo

Primer positivo: 15 de enero

WWTP1 (Besòs)



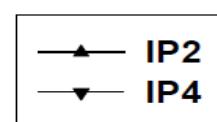
WWTP2 (El Prat)



Primer positivo: 15 de enero

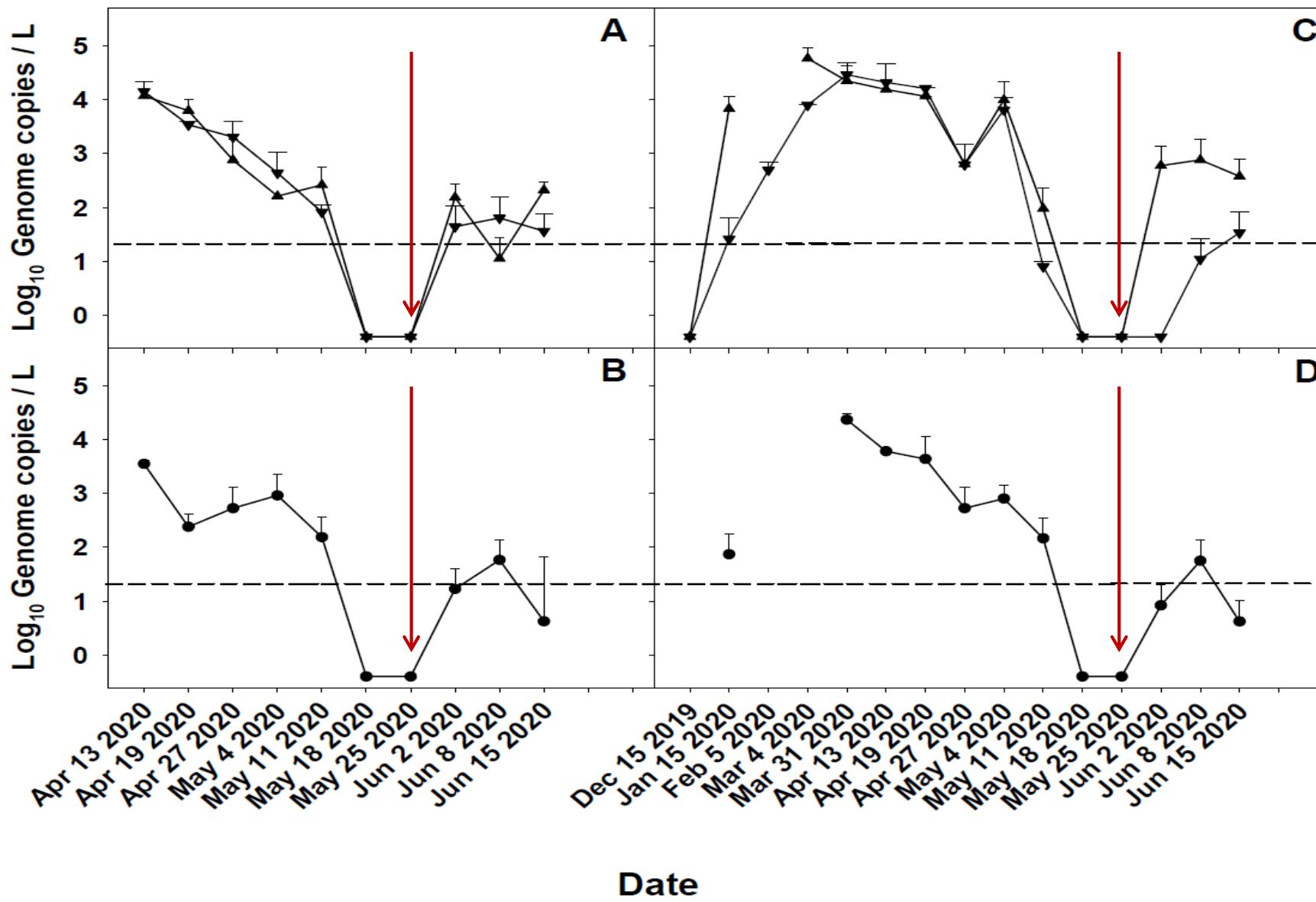
Primer caso COVID-19
confirmado: 25 de febrero

Date

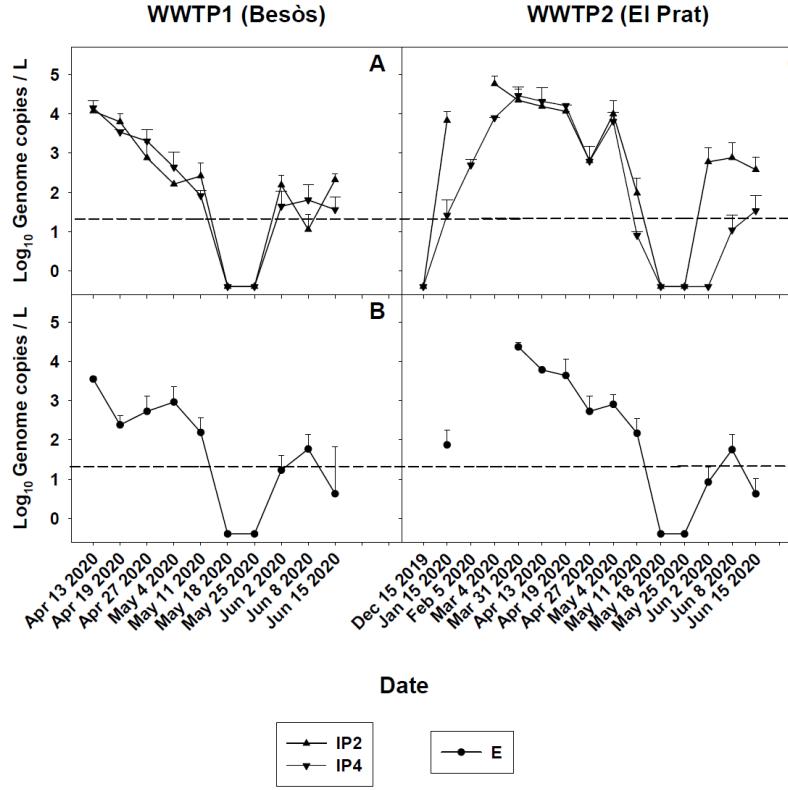


WWTP1 (Besòs)

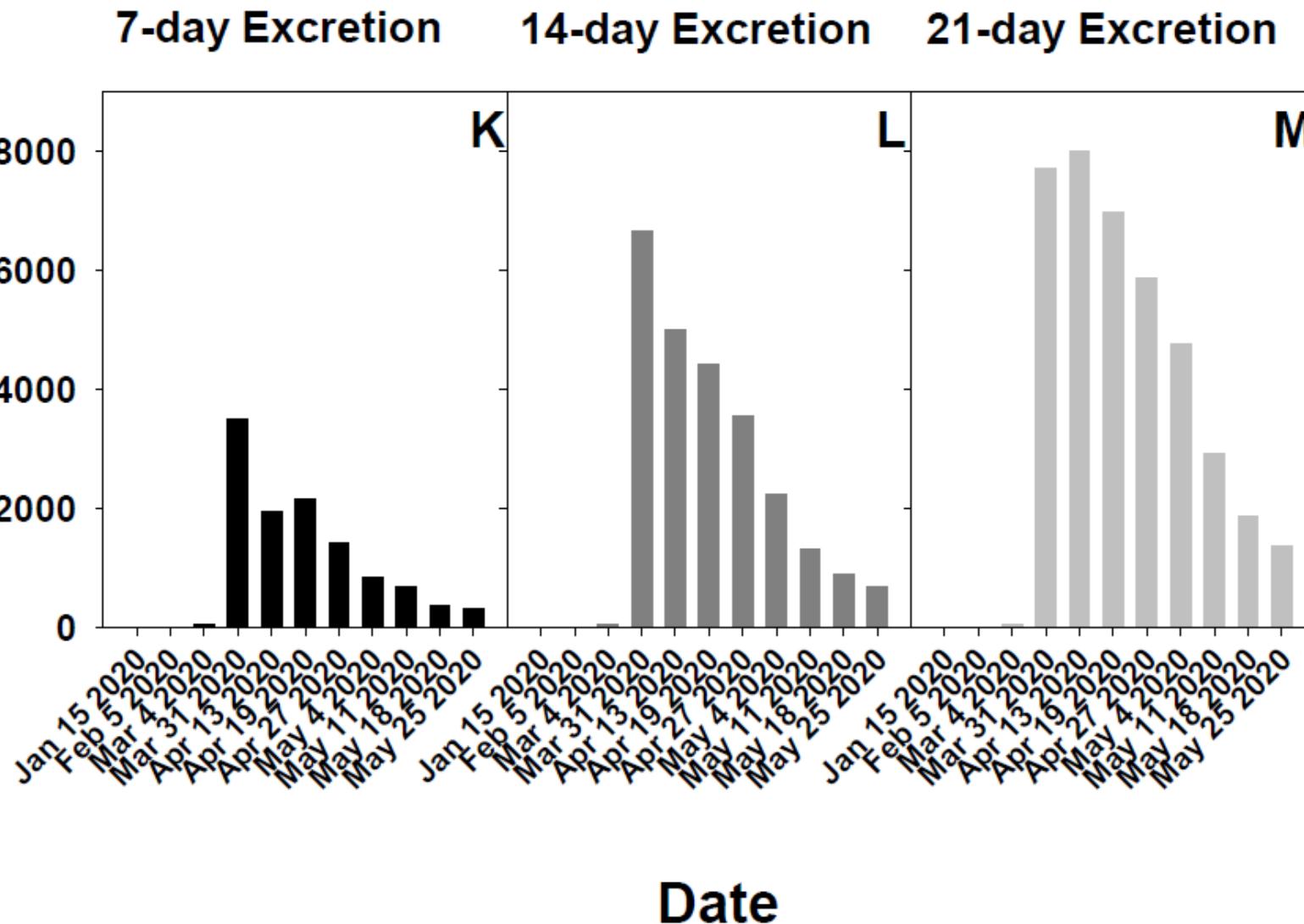
WWTP2 (El Prat)



Entrada Fase 1: 25 de Mayo



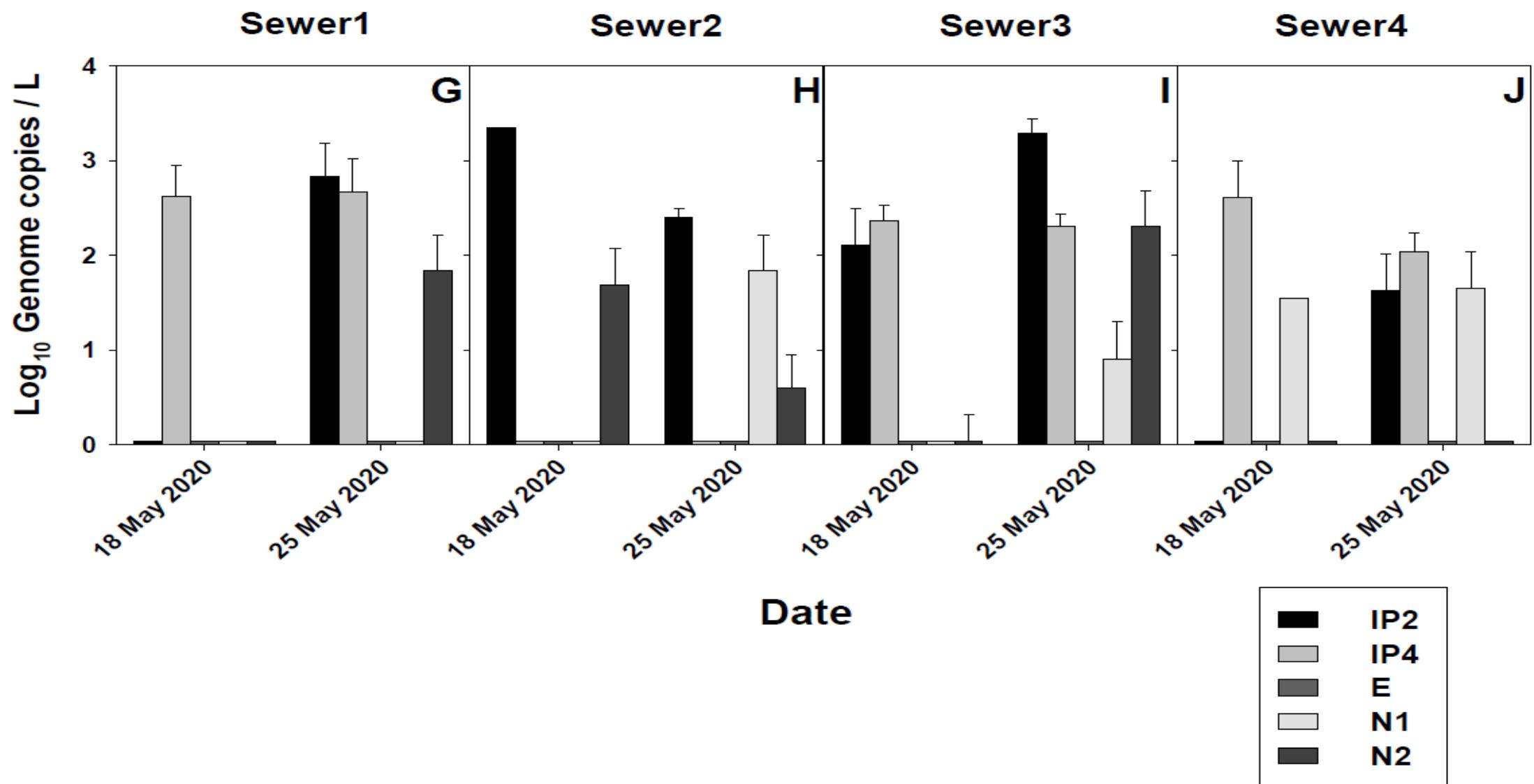
Cumulative shedders



VIRUS ENTÈRICS



SARS-CoV-2 in Barcelona sewers – End of May 2020



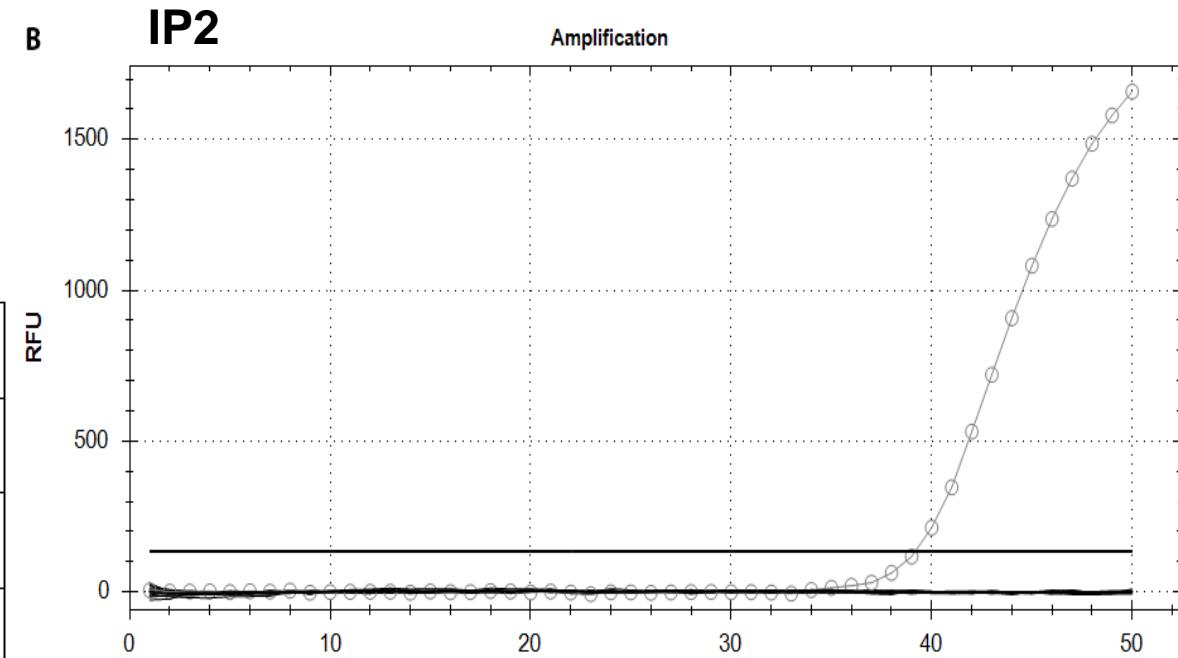
SARS-CoV-2 in raw sewage samples from March 2019

A

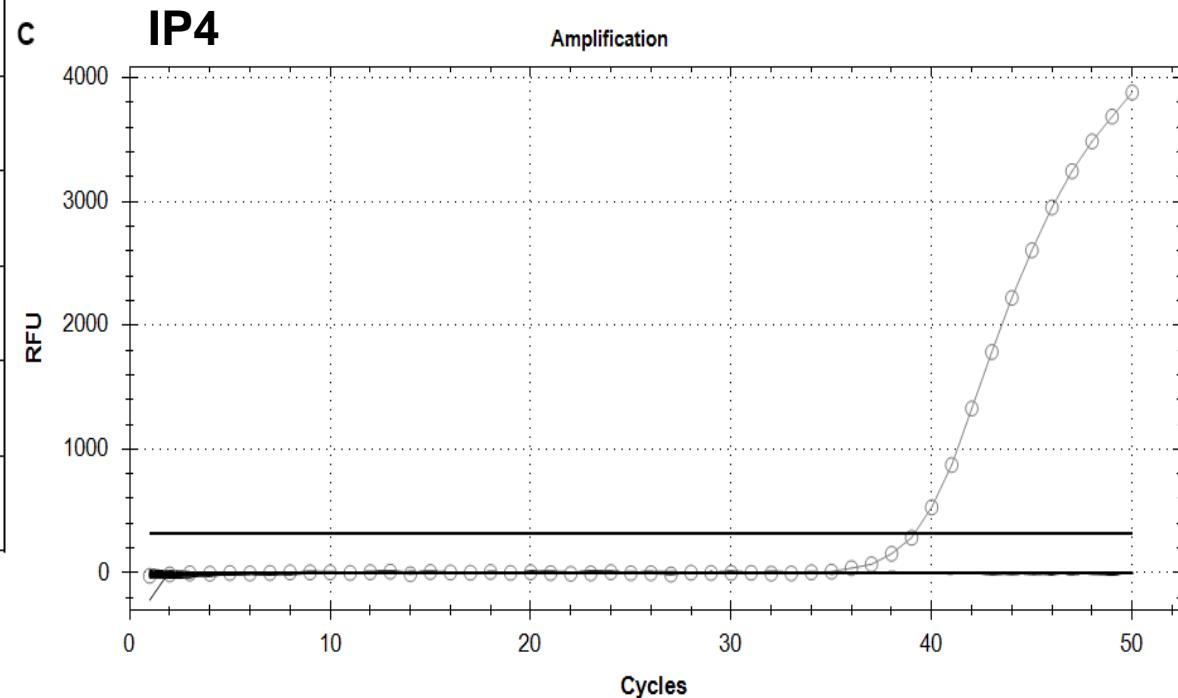
	IP2	IP4	E	N1	N2
January 16, 2018	No Ct	No Ct	No Ct	No Ct	No Ct
February 6, 2018	No Ct	No Ct	No Ct	No Ct	No Ct
March 6, 2018	No Ct	No Ct	No Ct	No Ct	No Ct
January 15, 2019	No Ct	No Ct	No Ct	No Ct	No Ct
March 12, 2019	6.4×10^2 *	8.3×10^2 *	No Ct	No Ct	No Ct
September 10, 2019	No Ct	No Ct	No Ct	No Ct	No Ct
October 2, 2019	No Ct	No Ct	No Ct	No Ct	No Ct
November 6, 2019	No Ct	No Ct	No Ct	No Ct	No Ct
December 11, 2019	No Ct	No Ct	No Ct	No Ct	No Ct

* Genome copies / L

B



C

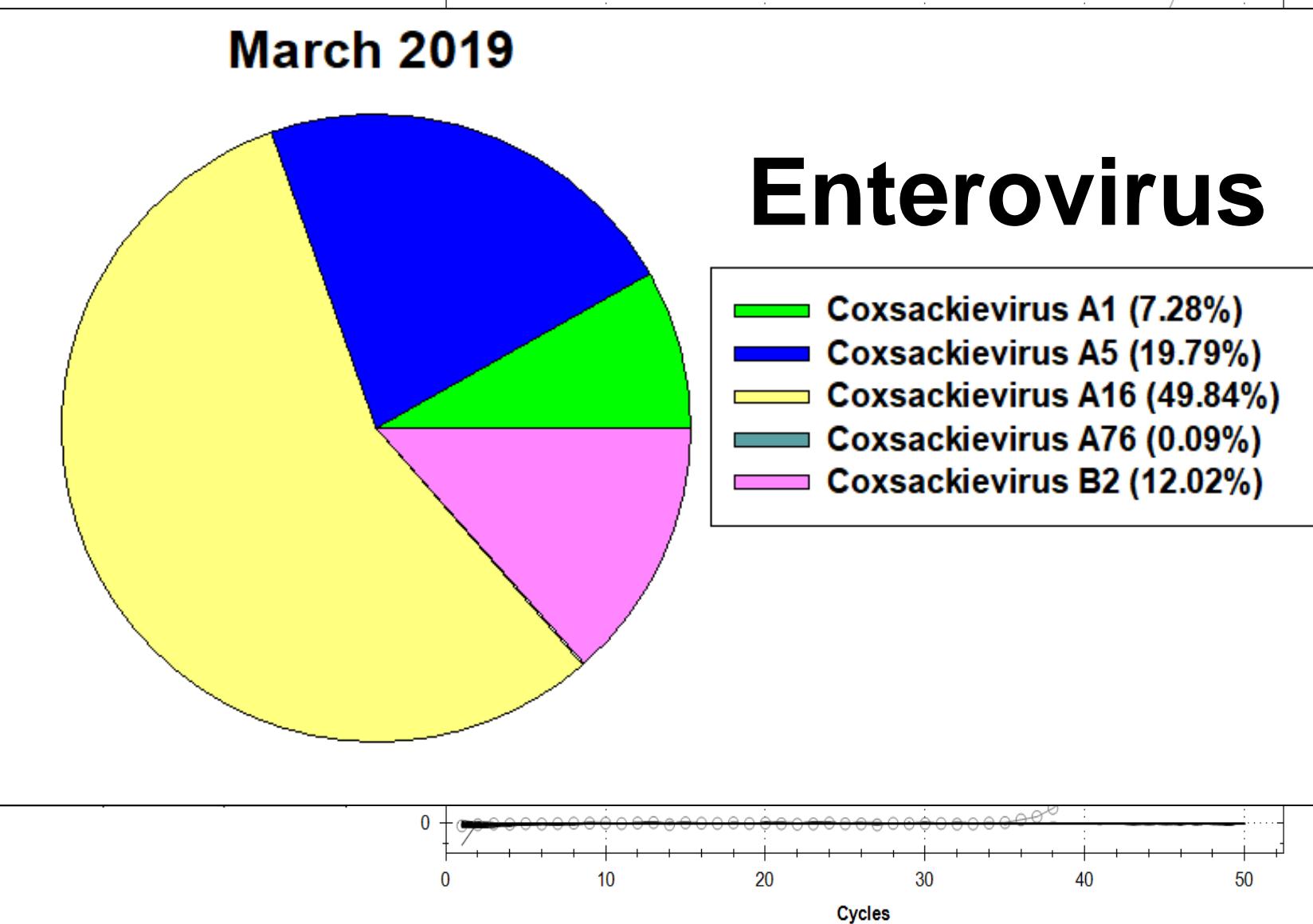


SARS-CoV-2 in raw sewage samples from March 2019

A

	IP2	IP4
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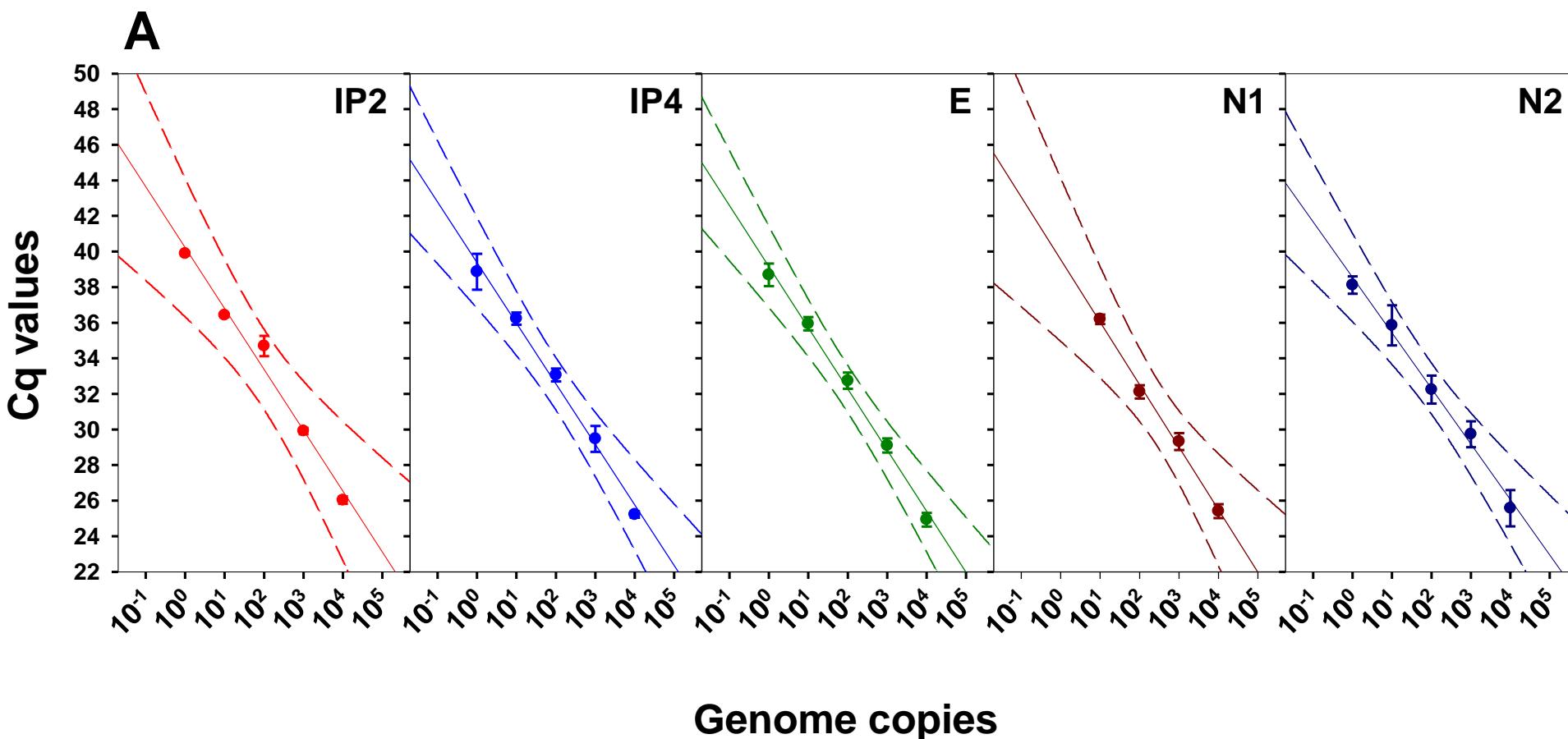
SARS-CoV-2 has been circulating in northern Italy since December 2019: evidence from environmental monitoring

Giuseppina La Rosa^{1*}, Pamela Mancini¹, Giusy Bonanno Ferraro¹, Carolina Veneri¹, Marcello Iaconelli¹, Lucia Bonadonna¹, Luca Lucentini¹, Elisabetta Suffredini²

¹ Department of Environment and Health, Istituto Superiore di Sanità, Rome, Italy

² Department of Food Safety, Nutrition and Veterinary Public Health, Istituto Superiore di Sanità, Rome, Italy

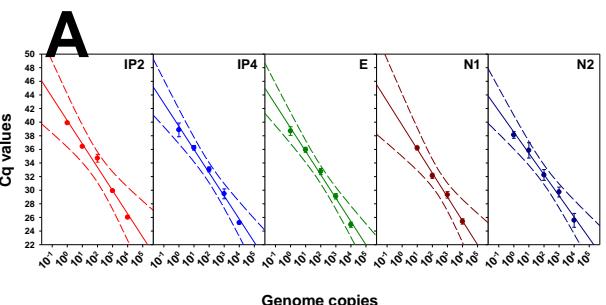
Average standard curves for each of the targets used in this study (IP2, IP4, E, N1 and N2). A. Regression line (solid lines) and their 99% confident (dashed lines). B. Parameters defining each of the curves.



B

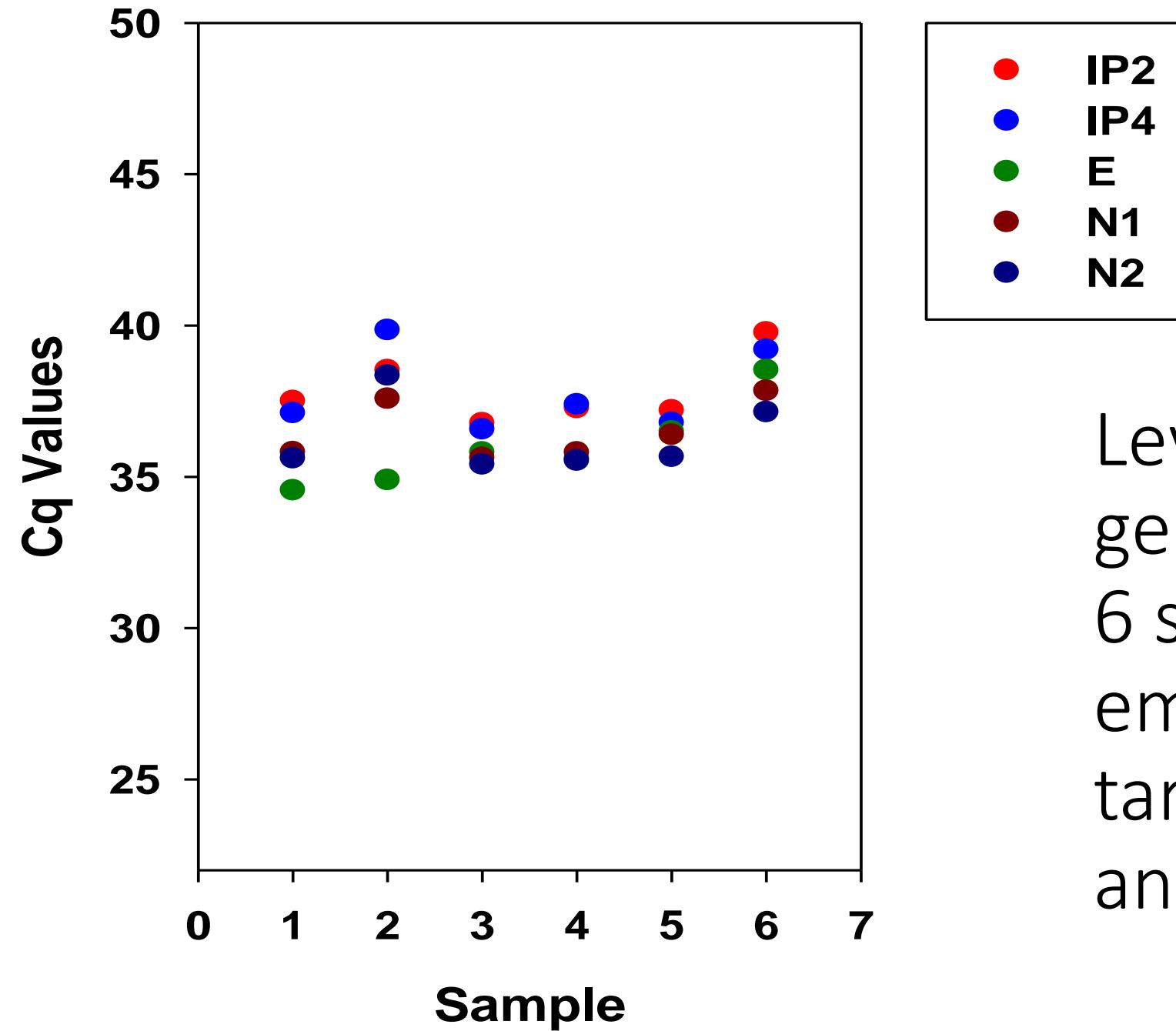
TARGET*	SLOPE	INTERCEPT	EFFICIENCY	R^2	Percent Replicates Positivity	
					10 ¹ Genome copies	10 ⁰ Genome copies
IP2	-3.421	40.228	96.0	0.981	33	33
IP4	-3.404	39.376	96.7	0.992	67	28
E	-3.436	39.152	95.5	0.994	100	50
N1	-3.513	39.543	92.6	0.995	100	0
N2	-3.120	38.540	109.2	0.991	100	67

Average standard curves for each of the targets used in this study (IP2, IP4, E, N1 and N2). A. Regression line (solid lines) and their 99% confident (dashed lines). B. Parameters defining each of the curves.



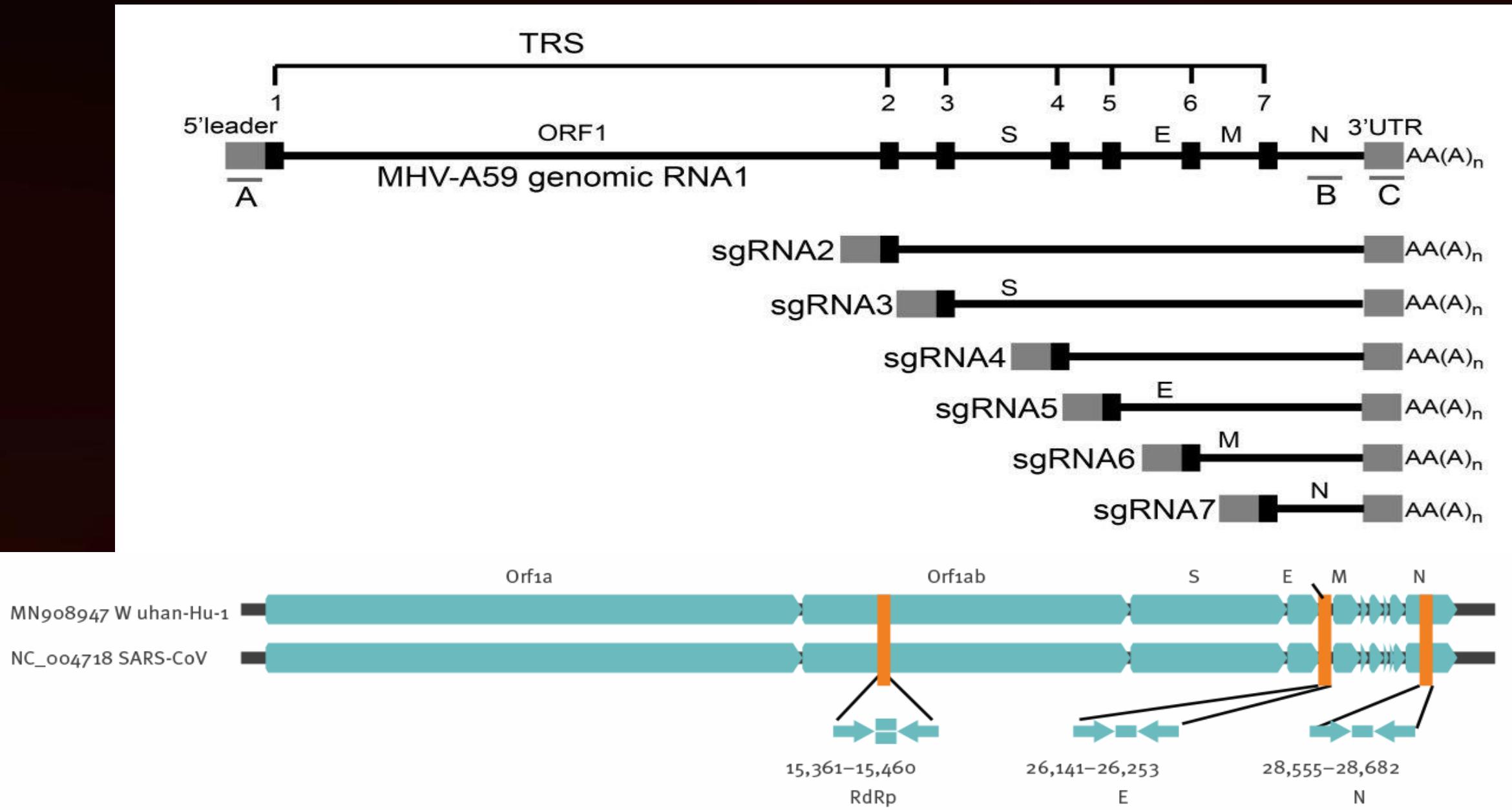
B

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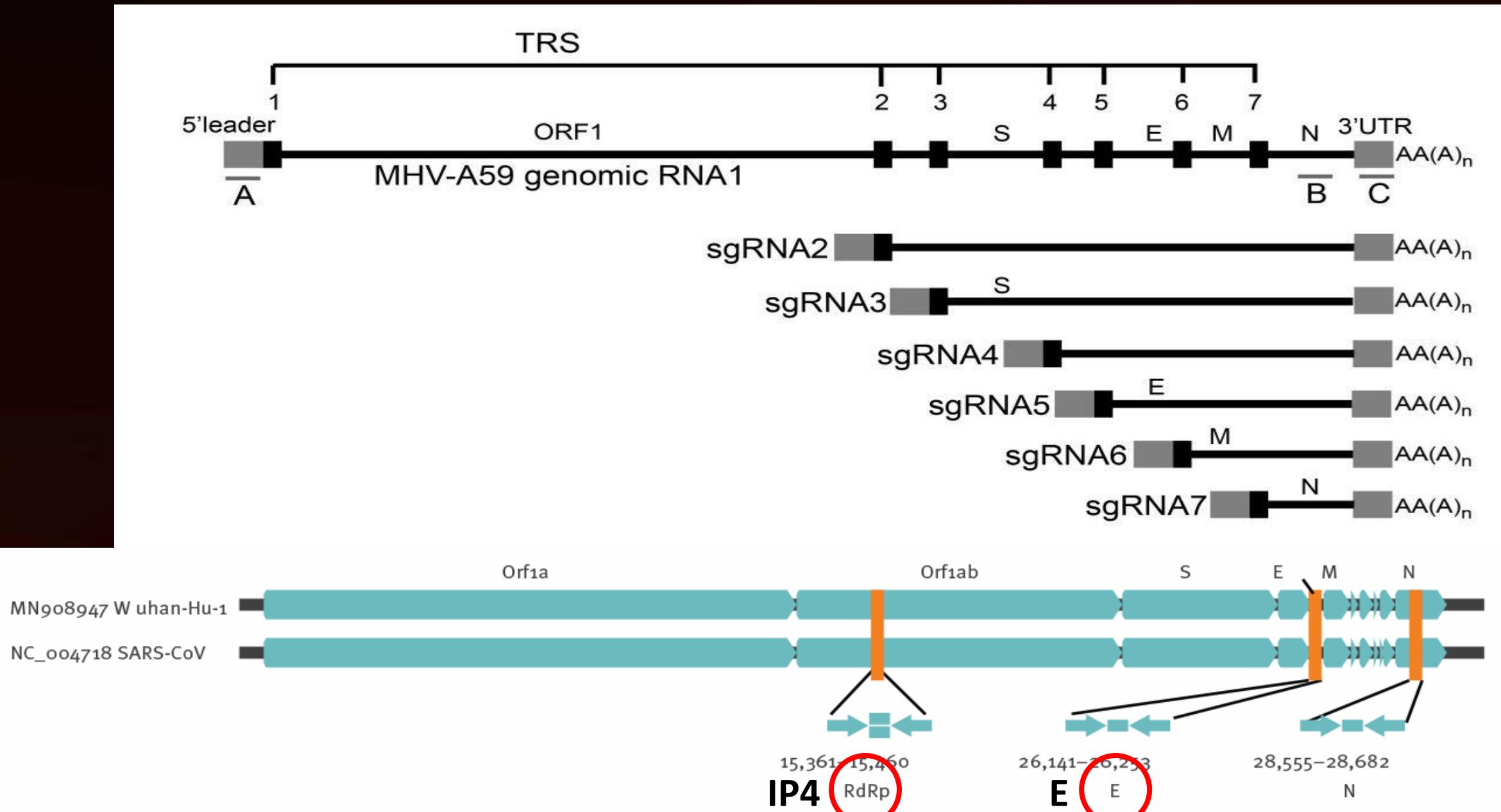


Levels of SARS-CoV-2 genomes (Cq Values) in 6 sewage samples employing the five targets (IP2, IP4, E, N1 and N2)

CoV Genome Organization and Expression



CoV Genome Organization and Expression



Actualizado: 02-05-2020 (Huesca, Teruel, Zaragoza),
28-04-2020 (A Coruña, Lugo, Ourense, Pontevedra),
01-05-2020 (Asturias, Baleares, Cantabria, Ceuta, La Rioja, Madrid, Melilla, Murcia, Navarra), 30-04-2020 (Resto)



La vigilancia del SARS-CoV-2 en aguas residuales es una herramienta de alerta rápida para la COVID-19

Fuentes: recopilación 'ProvidencialData19' de numeroteca (<https://github.com/monterea34/escovid19data>), INE (Padrón municipal a 1 de enero de 2019), gadm.org

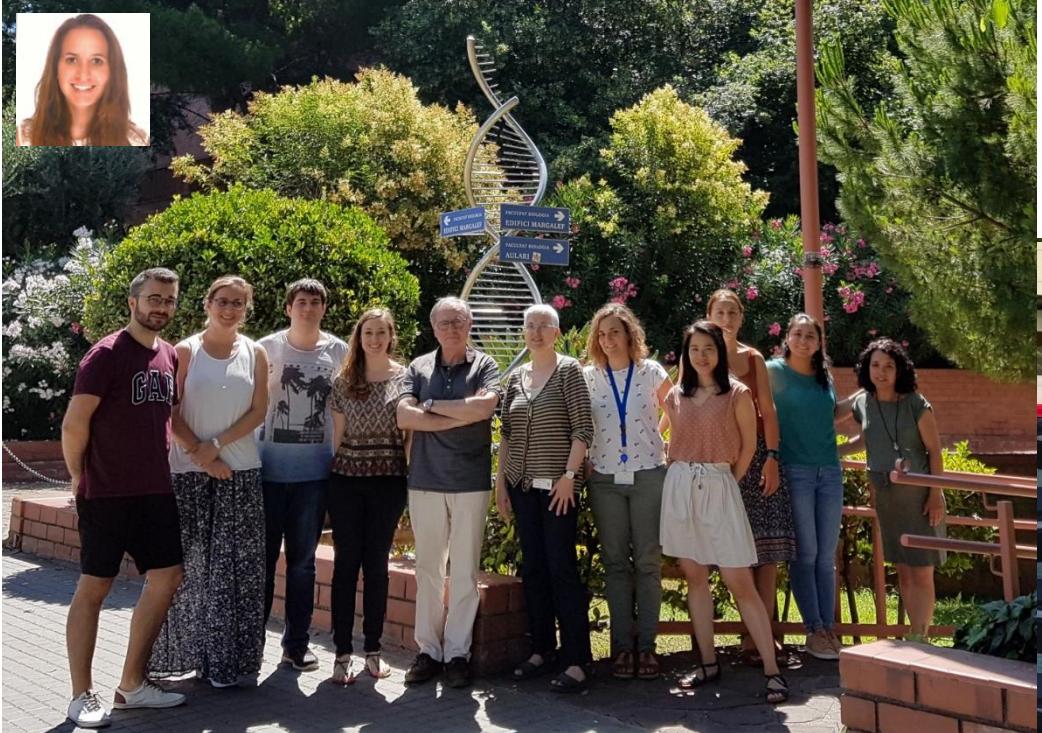
El coronavirus en España

146.690 casos confirmados



COVID-19 (02/05/20)





Análisis SARS-CoV-2 en aguas residuales

